

Interpretation: There are two processes governing the dark adaptation; one determining the first, fast phase, is strongly  $\text{Ca}^{2+}$ -dependent - probably controlled by the level of the intracellular  $\text{Ca}^{2+}$ -concentration-, the second one represents another slower recovery process which is much less  $\text{Ca}^{2+}$ -dependent.

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Interaction of specific neurotoxins with voltage-sensitive Na-channels in membrane vesicles

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The excitable face of the electroplaques in the electric organ of Electrophorus electricus propagates an action potential. The voltage-sensitive Na-channels in this membrane resemble pharmacologically those in nerve and striated muscle. To study their molecular and functional properties, Na-channel rich membrane fragments are prepared from the homogenized electric organ.

Standard preparation procedures including density gradient centrifugation yield fractions with specific tetrodotoxin binding up to 5 pMol/mg protein. After carrier-free column electrophoresis membranes with specific binding up to 30 pMol/mg are obtained, which corresponds to a tetrodotoxin-receptor density of ca 80/ $\mu\text{m}^2$ . A major part of the membrane fragments forms vesicles (electron microscopy), and at least 50% of them are inside-out (affinity chromatography on bound lectins / asymmetric binding of neurotoxins).

For binding studies, tetrodotoxin is chemically tritiated to high specific activity (with M.Rack, R.Stämpfli). It binds with a thermodynamic equilibrium constant  $K_D = (20 \pm 10) \text{ nM}$  to its receptor on these membrane fragments. The experimentally determined kinetic constants for a bimolecular binding equilibrium are  $k_f \gg 3 \times 10^5 \text{ M}^{-1} \text{ s}^{-1}$  and  $k_b = 2.5 \times 10^{-2} \text{ s}^{-1}$ .

To analyze the transport function of the channels, the efflux of  $^{22}\text{Na}$  from tetrodotoxin-receptor-rich membrane vesicles is monitored by filtration techniques. A modulation of the efflux by specific neurotoxins was observed only after having transiently established asymmetric Na- and K-ion concentrations inside and outside the vesicles. To improve the time resolution of these filtration assays, a technique of con-

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Extinction spectra of the membrane-bound chromoprotein show that at 0° C and pH 7.0 the system is stable and photoreversible for several days. The maximum of the extinction spectrum of ASTACUS rhodopsin (0° C, pH 7) is  $\lambda_{\text{max}} = 530 \text{ nm}$ . Rhodopsin (R) is isomerized nearly quantitatively to metarhodopsin (M) ( $\lambda_{\text{max}} = 500 \text{ nm}$ ) by irradiation at  $\lambda = 630\text{--}640 \text{ nm}$ .

$$R \begin{array}{c} \xrightarrow{\quad} M' \\ \xleftarrow{\quad} R' \end{array} \begin{array}{c} \xrightarrow{\quad} M \\ \xleftarrow{\quad} M' \end{array}$$

The activation energies were calculated from the slope of the Arrhenius plot and seem to be the same for both reactions,  $\Delta E_{M' \rightarrow M} = \Delta E_{R' \rightarrow R} = 22.5$  kcal mol<sup>-1</sup>.

Flash-spectroscopic measurements in the wavelength range from 430 to 650 nm show that the difference spectra of M-R' and M-R have the same maximum at 495 nm and the same minimum at 575 nm. This suggests that the absorption maxima of R and R' are similar, but differ in the molar extinction coefficient.

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